

CAPS Datasheets provide pest-specific information to support planning and completing early detection surveys.

Rhabdoscelus obscurus

Scientific Name

Rhabdoscelus obscurus (Boisduval, 1835)

Synonym(s):

Rhabdocnemis obscura (Boisduval, 1835)

Sphenophorus obscurus (Boisduval, 1835)

Calandra obscura Boisduval, 1835

Common Name

New Guinea sugarcane weevil, cane weevil borer

Type of Pest

Weevil, borer

Taxonomic Position

Class: Insecta, **Order:** Coleoptera, **Family:** Dryophthoridae

Pest Recognition

This section describes characteristics of the organism and symptoms that will help surveyors recognize possible infestations/infections in the field, select survey sites, and collect symptomatic material. For descriptions of diagnostic features, see the Identification/Diagnostic resources on the AMPS pest page on the CAPS Resource and Collaboration website.

Pest Description

Adults

Adults are approximately $\frac{1}{4}$ to $\frac{1}{2}$ inch long and $\frac{1}{8}$ to $\frac{1}{4}$ inch wide, with a curved, beak-like projection (rostrum) (Figs. 1, 2). Adults are dull, light brown with reddish brown to dark brown markings on the upper surface of the thorax and hardened outer wing covers (elytra) (Figs. 1, 2). These markings form a variable color pattern; six color patterns have been identified (Fig. 3). The elytra have long grooves running lengthwise along the back (Napometh et al., 1972; Terry, 1907). For additional information and images see the [CAPS Screening Aid](#) available for this pest. Adults are mostly active at night, hiding on host plants or in plant debris on the ground during the day (Napometh et al., 1972; Terry, 1907) and occasionally fly during the hottest part of the day (Koebele, 1900; Napometh et al., 1972).



Figure 1. New Guinea sugarcane weevil adult.
Photo Credit: G. Curt Fiedler, University of Guam.



Figure 2. Adult New Guinea sugarcane weevil: pinned specimen (**Top**) and live adult in *Pritchardia* spp. palm (**Bottom**). Photo credit: (**Top**) Aubrey Moore/Wikimedia Commons, [CC BY-SA 4.0](https://creativecommons.org/licenses/by-sa/4.0/); (**Bottom**) Whitney Cranshaw, Colorado State University, Bugwood.org, [CC BY-NC 3.0 US](https://creativecommons.org/licenses/by-nc/3.0/us/).

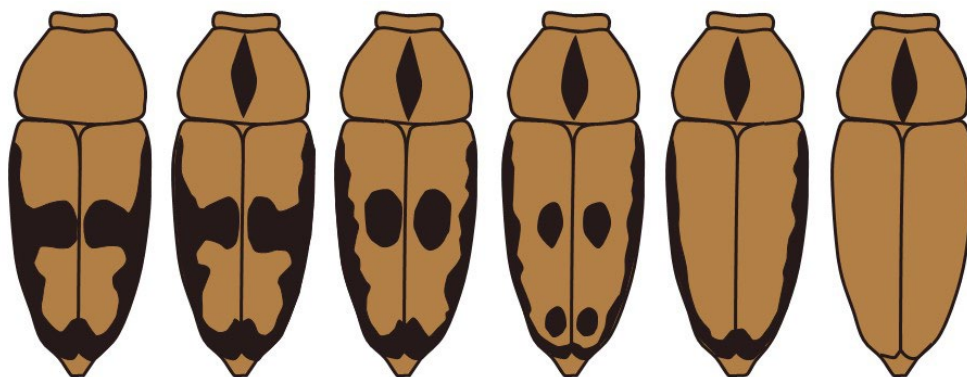


Figure 3. Weevil color variations on the thorax and elytra, illustrated after Fig. 19 in Napometh et al. (1972)

Eggs

Surveyors should not expect to identify eggs in the field. Eggs are small and oval, with a tough outer membrane. They are translucent white when laid but turn opaque white as they age (Napometh et al., 1972). Eggs are laid singly on host plants (Desmier De Chenon et al., 2001; Napometh et al., 1972).

Larvae

Larvae are approximately $\frac{1}{18}$ inch long in the first instar and $\frac{5}{8}$ inch long and $\frac{3}{8}$ inch wide when fully developed. The body of the larva is milky white and translucent, and the head is reddish orange with dark jaws (mandibles) (Fig. 4 Left). Larvae are wrinkled, legless, and covered with bristles (setae) (Muir and Swezey, 1916; Napometh et al., 1972; Terry, 1907). Larvae can be found in feeding galleries inside host plants, typically in the stalk or trunk (Desmier De Chenon et al., 2001; Dharmaraju et al., 1979; Napometh et al., 1972).



Figure 4. Larva inside a *Pritchardia* spp. palm (**left**) and a palm weevil cocoon (**right**). Photo credit: Whitney Cranshaw, Colorado State University, Bugwood.org, [CC BY-NC 3.0 US](https://creativecommons.org/licenses/by-nc/3.0/us/).

Pupae

Pupae are initially creamy or yellowish white and gradually darken to the colors of the future adult. Pupae are typically covered in cocoons made of fibers and frass (Fig. 4 Right) (Napometh et al., 1972; Terry, 1907). Pupation occurs in the feeding galleries created by the larvae (Desmier De Chenon et al., 2001; Napometh et al., 1972).

Signs and Symptoms

Sugarcane

Sugarcane plants infested with New Guinea sugarcane weevil may have round exit holes, frass-packed tunnels, visible feeding scars and cracks, and window-like openings in the cane (Naaz et al., 2020; Napometh et al., 1972; Samson et al., 2015).

Palm

Infested palm trees may have round exit holes, frass-packed tunnels, visible feeding scars and cracks, trunk staining, a jelly-like substance coming from holes in leaves or stems, and splitting near the base or further up the trunk (Fig. 6) (Dharmaraju et al., 1979; Halfpapp and Storey, 1991). In young palms, larvae may completely mine the center of stems and destroy the plants (Halfpapp and Storey, 1991). Adults may be found sheltering at the base of inflorescences or under leaf bases (Halfpapp and Storey, 1991). Cocoons are present within the trunks of dead plants or protruding through splits in the trunk (Halfpapp and Storey, 1991).



Figure 6. Symptoms of New Guinea sugarcane weevil infestation: exit hole wounds on *Pritchardia* palm trunk (**left**), close-up of exit holes (**top right**), frass and galleries caused by larval feeding (**bottom right**). Photo credit: Whitney Cranshaw, Colorado State University, Bugwood.org, [CC BY-NC 3.0 US](https://creativecommons.org/licenses/by-nc/3.0/us/).

Easily Mistaken Species

New Guinea sugarcane weevil can be confused with other species of *Rhabdoscelus*; however, none of these species are present in the United States. Other genera in the Dryophthoridae are similar in appearance to New Guinea sugarcane weevil, including *Rhynchophorus*, *Cosmopolites*, *Metamasius*, *Sphenophorus*, and *Scyphophorus* (Royals et al., 2017). Use the images and key available in the [CAPS Screening Aid](#) to help differentiate *Cosmopolites*, *Metamasius*, *Scyphophorus*, and *Sphenophorus* from

Rhabdoscelus. Additionally, Brodel (2002) is available on the AMPS page and provides a more technical key to domestic and intercepted Dryophthoridae.

Some *Rhynchophorus* species attack New Guinea sugarcane weevil hosts (i.e., palms and sugarcane), but *Rhynchophorus* are typically 2-2.5 times longer and 3 times wider than the New Guinea sugarcane weevil (Hoddle et al., 2024; Royals et al., 2017).

Rhynchophorus cruentatus infests various palm species, especially *Sabal palmetto* (cabbage palmetto) and *Serenoa repens* (saw palmetto) (Weissling and Giblin-Davis, 2023). It is native to Florida and the southeastern United States (Fig. 7a) (Hunsberger et al., 2000; Thomas, 2010).

Rhynchophorus palmarum infests various palm species, including *Cocos nucifera* (coconut), *Elaeis guineensis* (African oil palm), and *Phoenix spp.* (date palm), as well as *Saccharum officinarum* (sugarcane) (de la Torre et al., 2010). *Rhynchophorus palmarum* is established in California and has been detected in Arizona and Texas (Fig. 7b) (Hoddle et al., 2021; NAPPO, 2015; Villanueva and Esparza-Díaz, 2013).

Rhynchophorus vulneratus (previously believed to be *R. ferrugineus*) is a damaging palm pest and was detected and eradicated in California (Rugman-Jones et al., 2013).

Rhynchophorus ferrugineus, also a damaging palm pest, is not known to be present in the United States, but is present in the Caribbean (Fig. 7c) (Roda et al., 2011).

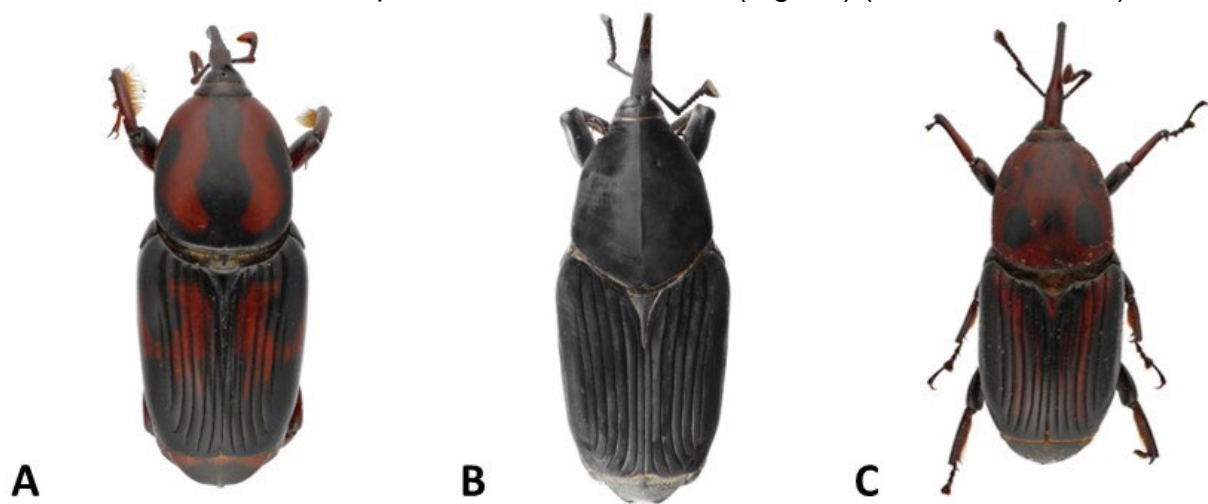


Figure 7. *Rhynchophorus* weevils that use similar hosts as New Guinea sugarcane weevil: *R. cruentatus* (A), *R. palmarum* (B), and *R. ferrugineus* (C). Photo credit: Hanna Royals, Screening Aids, USDA APHIS PPQ, Bugwood.org; [CC BY-NC 3.0 US](https://creativecommons.org/licenses/by-nc/3.0/us/).

The *Metamasius hemipterus* complex are pests of sugarcane, palms, and tropical fruits. *Metamasius sericeus* (= *M. hemipterus sericeus*) has become established in Florida and records from California and Mississippi are also likely *M. sericeus* (Fig. 8a) (CDFA, 2016; Palmieri et al., 2022; PPQ, 2025; Sosa Jr. et al., 1997; Thorn et al., 2019). *Metamasius hemipterus* (= *M. hemipterus hemipterus*) is present in Puerto Rico and recently became established in Hawaii (PPQ, 2025).

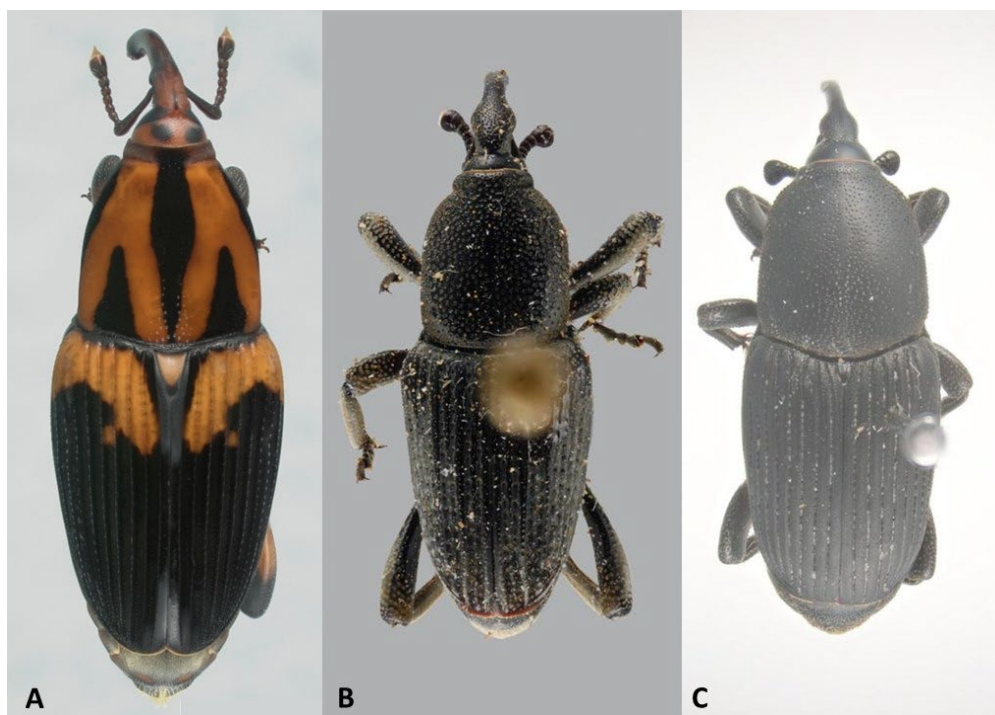


Figure 8. Other Dryophthoridae weevils that are similar to New Guinea sugarcane weevil: *Metamasius hemipterus* (A), *Cosmopolites sordidus* (B), and *Scyphophorus acupunctatus* (C). Photo credits: (A) Natasha Wright, Braman Termite & Pest Elimination, Bugwood.org; (B) Jennifer C. Girón Duque, Museum of Texas Tech University, Bugwood.org; and (C) Pest and Diseases Image Library, Bugwood.org; all images [CC BY-NC 3.0 US](https://creativecommons.org/licenses/by-nc/3.0/us/).

Cosmopolites sordidus is a pest of banana and is present in Florida, Hawaii, and Puerto Rico (Fig. 8b) (Gettman et al., 1992; Pantoja et al., 2006; Woodruff and Fasulo, 2021). *Scyphophorus acupunctatus* is a pest of agave in the southern United States and Hawaii (Fig. 8c) (Vaurie, 1971). *Sphenophorus* is a genus with multiple species native to North America (Vaurie, 1967). However, only *S. levis* and *S. incurrens* are pests of sugarcane, and they are not present in the United States (Casteliani et al., 2020; Illescas-Riquelme et al., 2016).

Commonly Encountered Non-targets

The approved survey method for *R. obscurus* is a bucket trap with a lure blend (2-methyl-4-octanol and ethyl acetate) and split sugarcane as food bait. The lure components may attract beetles and other insects including caddisflies, flies, springtails, stink bugs, and parasitoid wasps (Baraldi et al., 2002; El-Sayed, 2025).

Potential beetle non-targets that are attracted to one or more components of the lure blend and that are present in the United States include the easily mistaken species *Metamasius hemipterus* (= *M. hemipterus hemipterus*), *M. sericeus* (= *M. hemipterus sericeus*), *Rhynchophorus cruentatus*, and *R. palmarum* as well as *Carpophilus freemani*, *C. hemipterus*, *C. lugubris*, *C. mutilatus*, *Glischrochilus fasciatus*, *G. quadrisignatus*, and the genus *Phyllophaga* (Bartelt and Hossain, 2010; Bartelt et al., 1994; Giblin-Davis et al., 1994; Krell et al., 2015; Lin and Phelan, 1991; Milosavljević et al., 2019; Thorn et al., 2019).

Note that these species have not been verified to be attracted to *Rhabdoscelus obscurus* pheromone traps and that non-targets encountered during CAPS surveys will vary by region.

Biology and Ecology

Female weevils lay their eggs singly near internodes or leaf sheaths in sugarcane, in the trunk bark of young coconut palms, and on petioles or the base of fruit bunches in oil palms (Desmier De Chenon et al., 2001; Dharmaraju et al., 1979; Napometh et al., 1972). When laying eggs, females search for old holes, feeding scars, cracks from wind or rat damage, or they may chew shallow cavities in the host themselves (Muir and Swezey, 1916; Napometh et al., 1972). Egg development takes 3-7 days (Desmier De Chenon et al., 2001; Napometh et al., 1972; Terry, 1907).

Newly-hatched larvae bore downward into sugarcane stalks or coconut palm trunks, or into the petioles, rachises, and fruits of oil palms, creating galleries (feeding tunnels) that get larger as larvae develop (Desmier De Chenon et al., 2001; Dharmaraju et al., 1979; Napometh et al., 1972). Older larvae cut window-like openings in the sugarcane (Napometh et al., 1972). Larval development takes 45-82 days (Desmier De Chenon et al., 2001; Napometh et al., 1972).

Pre-pupae weave cocoons out of fibers and frass. Pupation takes place inside cocoons, which may occur in tunnels in sugarcane or coconut palms or amongst fruit bunches in oil palms (Desmier De Chenon et al., 2001; Napometh et al., 1972). The pre-pupal stage takes 6-9 days, then molts to become the pupa, which develops for approximately 7-12 days (Desmier De Chenon et al., 2001; Napometh et al., 1972).

After the adults emerge from the pupa, they remain inside the cocoon for an additional 10-14 days before digging their way out. Once active, adults feed, mate, and make short flights in the evening/night, with a smaller peak of activity in the morning. Weevils are attracted to volatiles emitted from injured or fermenting sugarcane and use these compounds to find hosts; excessive damage to sugarcane by rats, heavy winds, and severe weather events likely releases large amounts of these volatiles and is correlated with increased weevil damage (Muir and Swezey, 1916; Sallam et al., 2001). During the day, adults are less active, hiding in cracks or wounds on sugarcane stalks, between loosened sheaths of sugarcane leaves, or among plant debris on the ground (Napometh et al., 1972; Terry, 1907). Adults may infrequently fly during the hottest part of the day (Koebele, 1900; Napometh et al., 1972).

To find mates, weevils aggregate using a male pheromone that attracts both sexes (Chang and Curtis, 1972; Giblin-Davis et al., 2000). After mating, females lay most of their eggs in the first 16 weeks, with an average of 146 eggs over their lifetime. Their egg laying behavior is often irregular, alternating between periods of oviposition and periods with no egg-laying activity (Napometh et al., 1972). Adults are long-lived, surviving for at least 25 weeks in the field and up to 70 weeks in the laboratory (Napometh et al., 1972; Van Zwaluwenburg and Rosa, 1940).

New Guinea sugarcane weevil has 2-4 overlapping generations per year (Halfpapp and Storey, 1991; Napometh et al., 1972; Sallam et al., 2004). In Australia, the weevil is active throughout the year, with populations lowest in the winter and peaking in the summer to fall (Halfpapp and Storey, 1991; Sallam et al., 2004; Sallam et al., 2007). Weevil activity increases during higher temperatures (Sallam et al., 2004), but we could not find information on the minimum and maximum lethal temperatures for this pest. Areas with higher rainfall may be associated with higher weevil populations (Robertson and Webster, 1995; Tamanikaiyaroi, 1997).

Pathogens or Associated Organisms Vected

We found no evidence that New Guinea sugarcane weevil directly vectors any pathogens. However, feeding damage by the weevil facilitates the entry of secondary pathogens including *Colletotrichum falcatum*, a causative agent of red rot (Malathi et al., 2011; Tamanikaiyaroi, 1997).

Known Hosts

Sugarcane is the major host, and softer varieties are more heavily damaged (Muir and Swezey, 1916). The New Guinea sugarcane weevil also attacks a variety of palms and other occasional hosts (Halfpapp and Storey, 1991; Koebele, 1900; Muir and Swezey, 1916).

Minor hosts include corn, banana, and papaya (Muir and Swezey, 1916; Zimmerman, 1968). Although *R. obscurus* has been found on these hosts, we could find no evidence that these hosts are commonly infested.

The host list below includes cultivated and wild plants that 1) are infected or infested by the pest under natural conditions, 2) are frequently described as major, primary, or preferred hosts, and 3) have primary evidence for feeding and damage documented in the literature. Plants are highlighted in bold if they are commercially produced and the pest causes economically significant damage.

Table 1. Preferred hosts of *R. obscurus*.

Scientific Name	Common Name	Type/Use	References/Notes
<i>Aiphanes horrida</i> (= <i>Aiphanes caryotifolia</i>)	coyure palm	Cultivated	(Halfpapp and Storey, 1991)
<i>Archontophoenix alexandrae</i> *	Alexandra palm*	Cultivated	(Halfpapp and Storey, 1991)
<i>Archontophoenix cunninghamiana</i> *	Bangalow palm*	Cultivated	(Halfpapp and Storey, 1991)
<i>Areca catechu</i>*	betel-nut palm*	Cultivated	(Halfpapp and Storey, 1991; Reddy et al., 2011)
<i>Bactris gasipaes</i>	peach palm	Cultivated	(Halfpapp and Storey, 1991)
<i>Carpentaria acuminata</i> *	carpentaria palm*	Cultivated	(Halfpapp and Storey, 1991)
<i>Caryota mitis</i> *	clustered fishtail palm*	Cultivated	(Halfpapp and Storey, 1991)

Scientific Name	Common Name	Type/Use	References/Notes
<i>Caryota urens</i> *	fishtail palm*	Cultivated	(Koebele, 1900)
<i>Clinostigma savoryanum</i> (= <i>C. savoryana</i>)	Pacific beauty palm	Cultivated	(Karube et al., 2009)
<i>Cocos nucifera</i>*	coconut palm*	Cultivated	(Bianchi and Owen, 1965; Halfpapp and Storey, 1991)
<i>Dictyosperma album</i>	common princess palm	Cultivated	(Halfpapp and Storey, 1991)
<i>Dypsis decaryi</i> (= <i>Neodypsis decaryi</i>)	triangle palm	Cultivated	(Halfpapp and Storey, 1991)
<i>Dypsis lutescens</i> * (= <i>Chrysalidocarpus lutescens</i>)	golden-yellow palm*	Cultivated	(Halfpapp and Storey, 1991)
<i>Dypsis madagascariensis</i> (= <i>Chrysalidocarpus madagascariensis</i>)	green cane palm	Cultivated	(Halfpapp and Storey, 1991)
<i>Dypsis nodifera</i> (= <i>Phloga nodifera</i>)	NA	Cultivated	(Halfpapp and Storey, 1991)
<i>Dypsis</i> spp.*	butterfly palm*	Wild and cultivated	(Halfpapp and Storey, 1991)
<i>Elaeis guineensis</i> *	oil palm*	Cultivated	(Desmier De Chenon et al., 2001)
<i>Euterpe</i> spp.	euterpe	Wild and cultivated	(Halfpapp and Storey, 1991)
<i>Hyophorbe lagenicaulis</i> *	bottle palm*	Cultivated	(Halfpapp and Storey, 1991)
<i>Licuala</i> spp.	licuala	Wild and cultivated	(Halfpapp and Storey, 1991)
<i>Metroxylon sagu</i>	sago palm	Cultivated	(Muir and Swezey, 1916)
<i>Metroxylon salomonense</i>	Solomon's sago palm	Wild	(Halfpapp and Storey, 1991)
<i>Normanbya normanbyi</i>	black palm	Wild	(Halfpapp and Storey, 1991)
<i>Phoenix canariensis</i> *	Canary Island date palm*	Cultivated	(Halfpapp and Storey, 1991)
<i>Pigafetta filaris</i>	pigafetta palm	Cultivated	(Halfpapp and Storey, 1991)
<i>Pritchardia</i> spp.	loulou palm	Cultivated	(Conant et al., 2010; Koebele, 1900)
<i>Ptychosperma elegans</i> *	solitaire palm*	Cultivated	(Halfpapp and Storey, 1991)
<i>Roystonea regia</i> *	royal palm	Wild and cultivated	(Halfpapp and Storey, 1991)
<i>Saccharum officinarum</i>*	sugarcane*	Cultivated	(Naaz et al., 2020)
<i>Strelitzia reginae</i> *	bird-of-paradise*	Cultivated	(Pemberton, 1951)
<i>Syagrus romanzoffiana</i> *	queen palm*	Cultivated	(Halfpapp and Storey, 1991)
<i>Wodyetia bifurcata</i> *	foxtail palm*	Cultivated	(Halfpapp and Storey, 1991)

*Hosts with known US distribution (GBIF Secretariat, 2025; Kartesz, 2015; USDA-NRCS, 2025)

Known Distribution

Rhabdoscelus obscurus is native to New Guinea and surrounding islands (Halfpapp and Storey, 1991; Muir and Swezey, 1916). The weevil has been introduced to most islands in the Pacific, including Australia, Guam, Hawaii, Indonesia, and Japan (Desmier De Chenon et al., 2001; Muir and Swezey, 1916; Muniappan et al., 2004; Robertson et al., 1997; Takahashi, 1997).

Table 2. Countries where *R. obscurus* is known to occur.

Continent	Country	References/Notes
Asia	Indonesia	(Desmier De Chenon et al., 2001)
Asia	Japan	(Takahashi, 1997)
North America	United States (Hawaii)	(Giblin-Davis et al., 2000)
Oceania	Australia (incl. Christmas Island)	(Halfpapp and Storey, 1991; Muir and Swezey, 1916)
Oceania	Cook Islands	(Waterhouse, 1997)
Oceania	Federated States of Micronesia	(Bianchi and Owen, 1965)
Oceania	Fiji	(Singh et al., 2019)
Oceania	French Polynesia	(Zimmerman, 1968)
Oceania	Guam	(Bianchi and Owen, 1965)
Oceania	Marshall Islands	(Zimmerman, 1968)
Oceania	New Caledonia (France)	(Waterhouse, 1997)
Oceania	Niue	(Given, 1968)
Oceania	Northern Mariana Islands	(Bianchi and Owen, 1965)
Oceania	Palau	(Bianchi and Owen, 1965)
Oceania	Papua New Guinea	(Zimmerman, 1968)
Oceania	Samoa	(Dharmaraju et al., 1979)
Oceania	Solomon Islands	(Waterhouse, 1997)
Oceania	Tonga	(Cottrell-Dormer, 1941)
Oceania	Vanuatu	(Waterhouse, 1997)

Rhabdoscelus obscurus has been detected in and eradicated from the Netherlands (Plant Protection Service, 2009). There are also miscellaneous records for Malaysia, Taiwan, American Samoa, and Nauru (EPPO, 2025; Singh et al., 2019), but these could not be verified.

Status of infestation in the United States (September 2025)

Rhabdoscelus obscurus is present in Hawaii, Guam, and the Northern Mariana Islands, but has not been reported from the contiguous United States (Bianchi and Owen, 1965; Giblin-Davis et al., 2000).

Pest Importance

Larval feeding reduces sugarcane yield and quality and can cause mortality of young coconut and other palm plants (Halfpapp and Storey, 1991; Karube et al., 2009; Naaz et al., 2020; Reddy et al., 2011). In the early 1990s, the pest caused \$3.5 million in sugarcane losses in Queensland, Australia and has caused yield losses of up to 10% (Robertson and Webster, 1995; Zimmerman, 1968). Earlier reports in Hawaii state that over half the sugarcane crop was lost to this pest in some cases (Timberlake, 1926), suggesting that very high losses are possible in unmitigated conditions.

Rhabdoscelus obscurus is described as causing widespread or complete yield losses in nursery plants and betel nuts in Micronesia in the absence of effective controls (Reddy et al., 2011). Larvae bore near the growing tip of betel nut, which can kill the tree (Schreiner, 2000). During an outbreak of *R. obscurus* in Guam, 100% of coconut trees were damaged and no fruits were produced over several months (Bianchi and Owen, 1965). Severe weather and disease may have contributed to the severity of damage observed, though *R. obscurus* was thought to cause most of the damage (Bianchi and Owen, 1965).

Though the impact of *R. obscurus* to other palm species has not been quantified, palm growers have reported problems with *R. obscurus* ranging from mild to severe, and the palm genera described as most susceptible include *Carpentaria*, *Dypsis*, *Roystonea*, and *Wodyetia* (Halfpapp and Storey, 1991).

In 2023, sugarcane was grown commercially in Florida, Louisiana, and Texas, with an estimated harvest value of \$2 billion (NASS, 2025). Coconut, betel nut, and oil palms have very limited production in the continental United States (Broschat and Crane, 2020; NASS, 2025; Reynolds, 2015), but palms for ornamental and landscaping purposes are widespread in the United States (NASS, 2025; USDA-FAS, 2025).

Rhabdoscelus obscurus is listed as a harmful organism by Argentina, Brazil, China, Colombia, Ecuador, El Salvador, Honduras, India, Japan, Mexico, Paraguay, Peru, South Korea, Uruguay, and Venezuela (USDA-APHIS-PPQ, 2025). There may be trade implications with these countries if *R. obscurus* becomes established in the continental United States.

Pathway

New Guinea sugarcane weevils can spread through the movement of infested materials, including sugarcane plants and cut pieces used for propagation, nursery plants (i.e., ornamental palms), and cut palm trees (Gao et al., 2007; Halfpapp and Storey, 1991). We queried USDA's Agricultural Risk Management (ARM) system for "*Rhabdoscelus obscurus*" and "*Rhabdoscelus* spp." interceptions at U.S. ports of entry. We found it was detected over two dozen times, primarily in airport baggage from passengers departing Hawaii who were carrying sugarcane or coconut cuttings (USDA ARM, 2024), indicating a possible pathway.

Some natural dispersal is possible. Adults take short, erratic flights, infesting nearby cane fields. In a mark-release-recapture experiment, marked weevils were found up to ~1/3 mile away within three months (Van Zwaluwenburg and Rosa, 1940).

Use the PPQ Commodity Import and Export manual listed below to determine 1) if host plants or material are allowed to enter the United States from countries where the organism is present and 2) what phytosanitary measures (e.g., inspections, phytosanitary certificates, post entry quarantines, mandatory treatments) are in use. This manual is updated regularly.

Agricultural Commodity Import Requirements(ACIR) manual: ACIR provides a single source to search for and retrieve entry requirements for imported commodities.

Potential Distribution within the United States

Rhabdoscelus obscurus has been reported from areas corresponding to plant hardiness zones 10 and higher (Halfpapp and Storey, 1991; Sallam et al., 2007; Takeuchi et al., 2018). Due to suitable climate and ongoing sugar production (NASS, 2025; Sterk, 2024; Takeuchi et al., 2018), sugarcane production in Florida, Louisiana, Puerto Rico, and the U.S. Virgin Islands would be at risk if *R. obscurus* becomes established in the United States. Texas sugarcane production may also be at risk, though its last sugarcane grower and processor recently closed (Sterk, 2024). Florida, California, and Texas would be at risk due to their large inventory of palm plants for landscaping (NASS, 2025; Takeuchi et al., 2018). Additionally, since *R. obscurus* has been observed infesting palm plants in greenhouse conditions (Plant Protection Service, 2009), additional states with greenhouse production may be at risk in the United States.

Survey and Key Diagnostics

Approved Methods for Pest Surveillance*:

For the current approved methods and guidance for survey and identification, see Approved Methods for Pest Surveillance (AMPS) pest page on the CAPS Resource and Collaboration website, at <https://approvedmethods.ceris.purdue.edu/>.

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Versions

September 2013: Datasheet completed (Version 1)

July 2016: Minor revision (Version 1.1)

- NAPPFAST map removed

December 2025: Datasheet revised (Version 2)

- Added **Pest Recognition** section
- Added **Easily Mistaken Species** section
- Added **Commonly Encountered Non-targets** section
- Removed **Damage** section
- Updated **Synonyms**
- Updated **Pathogens or Associated Organisms Vectored** section
- Updated **Known Hosts** section
- Updated **Known Distribution** section
- Updated **Pest Importance** section
- Updated **Pathway** section
- Updated **Potential Distribution in the United States** section
- Updated guidance for **Approved Methods** section